

# CALCIUM AND MAGNESIUM SILICATE AND SOIL AS ENVIRONMENTAL STABILIZERS IN THE CULTIVATION OF NILE TILAPIA LARVAE IN THE RECIRCULATION SYSTEM

## *SILICATO DE CÁLCIO E MAGNÉSIO E SOLO COMO ESTABILIZADORES AMBIENTAIS NO CULTIVO DE LARVAS DE TILÁPIA-DO-NILO EM SISTEMA DE RECIRCULAÇÃO*

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### ABSTRACT

With the intensification of production systems and the need to reduce water use, there is a greater likelihood of sudden changes in water quality, leading to fish mortality. Therefore, it is necessary to develop techniques that increase environmental homeostasis. This experiment evaluated the influence of calcium-magnesium silicate and soil compared to calcium carbonate and calcium-magnesium carbonate, traditional alkalizers, as environmental stabilizers in cultivating Nile tilapia larvae in a recirculating system. The experiment was composed of five treatments: Control) aquarium containing only water; Soil) water and soil; Calcitic) water and calcium carbonate ( $\text{CaCO}_3$ ); Dolomite) water and dolomitic limestone ( $\text{CaCO}_3$   $\text{MgCO}_3$ ) and Silicate) water and calcium-magnesium silicate ( $\text{CaSiO}_3$   $\text{MgSiO}_3$ ). After 30 days, the performance parameters, weight, standard length, total length, Fulton condition factor, weight gain, biomass, biomass gain, and survival, did not differ among treatments. Among the water quality parameters, temperature (30 °C) and oxygen (5 to 6 mg L<sup>-1</sup>) were maintained with heaters and aeration throughout the period. The parameters conductivity, pH, redox potential, salinity, turbidity, ammonia, nitrite, nitrate, alkalinity, hardness, calcium, magnesium, and silica were measured. The pH was higher in the silicate and calcitic treatments than in the control. The dolomitic treatment was higher for the redox potential than in the soil. The electrical conductivity was higher in the soil treatment than in the control. The turbidity in the soil was higher than in the other treatments. Salinity was higher in treatments that received liming products but with low values. Ammonia concentration was higher in the control treatment than in the silicate. Nitrite and nitrate concentrations did not differ between treatments. Alkalinity was higher in the silicate treatment. Hardness was higher in the calcitic, dolomitic, and silicate treatments than in control and soil. Calcium dissolved in water was higher in the calcitic and silicate treatments than in the control and soil. Silica dissolved in water was higher in silicate. Calcium-magnesium silicate is a viable and recommended alternative for liming, as it presents results equivalent to calcitic and dolomitic limestone, traditional salts for this practice, which confirms its importance in maintaining water quality and fish performance. Although the soil has a buffering capacity in the water, it is less effective than other products and deserves further study.

**KEYWORDS:** Alkalinity, Hardness, Liming, pH, Water homeostasis.

## RESUMO

Com a intensificação dos sistemas de produção e a necessidade de redução do uso de água, há maior probabilidade de mudanças bruscas na qualidade da água, levando à mortalidade de peixes. Portanto, é necessário desenvolver técnicas que aumentem a homeostase ambiental. Este experimento avaliou a influência do silicato de cálcio-magnésio e do solo em comparação ao carbonato de cálcio e ao carbonato de cálcio-magnésio, alcalinizantes tradicionais, como estabilizadores ambientais no cultivo de larvas de tilápia do Nilo em sistema de recirculação. O experimento foi composto por cinco tratamentos: Controle) aquário contendo apenas água; Solo) água e solo; Calcítico) água e carbonato de cálcio ( $\text{CaCO}_3$ ); Dolomítico) água e calcário dolomítico ( $\text{CaCO}_3$   $\text{MgCO}_3$ ) e Silicato) água e silicato de cálcio-magnésio ( $\text{CaSiO}_3$   $\text{MgSiO}_3$ ). Após 30 dias, os parâmetros de desempenho, peso, comprimento padrão, comprimento total, fator de condição de Fulton, ganho de peso, biomassa, ganho de biomassa e sobrevivência, não diferiram entre os tratamentos. Entre os parâmetros de qualidade da água, a temperatura (30 °C) e oxigênio (5 a 6 mg  $\text{L}^{-1}$ ) foram mantidos com aquecedores e aeração durante todo o período. Os parâmetros condutividade, pH, potencial redox, salinidade, turbidez, amônia, nitrito, nitrato, alcalinidade, dureza, cálcio, magnésio e sílica foram medidos. O pH foi maior nos tratamentos silicato e calcítico do que no controle. O tratamento dolomítico foi maior para o potencial redox do que no solo. A condutividade elétrica foi maior no tratamento solo do que no controle. A turbidez no solo foi maior do que nos outros tratamentos. A salinidade foi maior nos tratamentos que receberam produtos de calagem, mas com valores baixos. A concentração de amônia foi maior no tratamento controle do que no silicato. As concentrações de nitrito e nitrato não diferiram entre os tratamentos. A alcalinidade foi maior no tratamento silicato. A dureza foi maior nos tratamentos calcítico, dolomítico e silicato do que no controle e solo. O cálcio dissolvido em água foi maior nos tratamentos calcítico e silicato do que no controle e solo. A sílica dissolvida em água foi maior no silicato. O silicato de cálcio-magnésio é uma alternativa viável e recomendada para calagem, pois apresenta resultados equivalentes aos calcários calcítico e dolomítico, sais tradicionais para essa prática, o que confirma sua importância na manutenção da qualidade da água e no desempenho dos peixes. Embora o solo tenha capacidade tampão na água, ele é menos eficaz que outros produtos e merece estudos mais aprofundados.

**PALAVRAS-CHAVE:** Alcalinidade; Calagem; Dureza; Homeostase da água, pH.

## INTRODUCTION

The annual production growth recorded since 2000 is due to the intensification process, where production management, food, and biosecurity have improved. In this sense, aquaculture needs to pay greater attention to the health of the production environment, where keeping water quality is a challenge<sup>1</sup>. Meeting the global demand for aquatic products and, at the same time, maintaining sustainability is a critical challenge where it is necessary to combine technological solutions, innovations, and best management practices, increasing efficiency in an ecosystem approach to aquaculture resources<sup>2</sup>.

Elevated alkalinity levels have been used in aquaculture to prevent rapid pH fluctuations, improve water quality, maintain the health and well-being of fish, and increase production in intensive systems characterized by numerous physical, chemical, and biological interactions<sup>3</sup>.

Although alkalinity, as well as total hardness, another environmental stability parameter closely related to alkalinity, are familiar variables in aquatic animal production, where both scientists and aquaculturists have some knowledge of methods for adjusting their concentrations, their chemical relationships and biological effects are more complex than generally realized or described in the literature<sup>4</sup>.

Alkalinity is necessary for pH stability, which has a direct impact on the toxicity of ammonia, CO<sub>2</sub>, and other metabolic compounds, as well as providing inorganic carbon for nitrifying bacteria, which are essential for the functioning of the culture, as well as on the efficiency of critical processes such as nitrification and CO<sub>2</sub> removal (Jafari et al., 2024)<sup>3</sup>. Interactions are diverse, with different responses requiring detailed assessments<sup>3-5</sup>.

Calcitic limestone, composed of calcium carbonate, and dolomitic limestone, calcium carbonate with magnesium, are the alkalizing agent generally used because they are easily accessible and cheap<sup>6</sup>. Other products that increase alkalinity have been used, such as calcium silicate, which is occasionally used<sup>4</sup>, sodium carbonate, which is highly soluble and reacts quickly been is safe for fish and humans as well as sodium bicarbonate and calcium hydroxide, this which, despite being dangerous, is commonly used because it is cheaper and more readily available than sodium carbonate<sup>7</sup>. However, with intensification, production in closed systems requires adjustments, such as using new products or the same alkalizing products with different quality and concentrations than those usually used<sup>5</sup>.

In the aquaculture intensive, the alkalizing agents also have been selected based on results associated with ease of obtaining and price, and this is the case with sodium silicate, sodium bicarbonate, and calcium carbonate powder used in the recirculation system<sup>8</sup>, and sodium bicarbonate, calcium carbonate or calcium hydroxide in a biofloc system<sup>9</sup>.

Sodium bicarbonate (NaHCO<sub>3</sub>) has been used in intensive systems due to its efficiency in rapidly increasing alkalinity and pH, being safe for both animals and fish farmers, in addition to being affordable<sup>8,9</sup>. Calcium hydroxide (Ca(OH)<sub>2</sub>) is also very efficient in increasing pH and alkalinity, producing good results in intensive production systems. However, because it is highly reactive, it can cause fish mortality and should be used more sparingly<sup>9</sup>. Calcium carbonate powder

increases pH and alkalinity for a prolonged period, taking longer to dissolve<sup>8</sup>, and generates greater turbidity, which can compromise fish growth<sup>9</sup>. Sodium silicate dissolves quickly, promptly altering the pH and presenting results closed with sodium bicarbonate and calcium carbonate<sup>8</sup>.

Although calcium silicate raises alkalinity and pH to a level slightly below that of agricultural limestone and has a water solubility similar to that of calcium carbonate<sup>10</sup>, there is rarely a study evaluating its efficiency in an intensive fish culture system, even in a conventional system, which generates a demand for studies with this product.

The sediment at the bottom of the tank is an essential part of the farming ecosystem, exchanging nutrients at the soil-water interface, with feldspars composed of calcium silicate raises the pH<sup>4</sup> while the aluminum acidifying<sup>10</sup> influencing water quality and, consequently, affecting animal growth and welfare<sup>11,12</sup>. Despite the importance of sediment in this ecosystem, sediment has yet to be studied as a possible factor in improving conditions in intensive systems of tilapia larva culture.

For the reasons described above, this work aimed to evaluate the influence of calcium-magnesium silicate and soil compared to traditional liming calcium carbonate and calcium and magnesium carbonate on tilapia larva culture.

## **MATERIAL AND METHODS**

The experiment to evaluate the effect of different forms of liming on Nile tilapia larvae was carried out during 30 days in the Aquaculture Laboratory of Aquatic Ecology, DZO - UFVJM, in Diamantina (Latitude 18°14'17 "South, longitude 43°36'40"West), located in the region of the Southern Espinhaço Ridge. The study was approved by the Ethics Committee on Animal Use (CEUA) of UFVJM (n° 031/2019 /CEUA-UFVJM) protocolo 001\2017. The experiment was composed of five treatments being: Control) aquarium containing only water; Soil) water and soil; Calcitic) water and calcium carbonate ( $\text{CaCO}_3$ ); Dolomite) water and dolomitic limestone (70%  $\text{CaCO}_3$  • 30%  $\text{MgCO}_3$ ) and Silicate) water and calcium-magnesium silicate ( $\text{CaSiO}_3$   $\text{MgSiO}_3$ ). The treatments were randomly distributed in a completely randomized design, with five replicates each, being 25 aquariums. The aquarium had 10 L, aeration ( $40 \text{ ml min}^{-1}$ ,  $>4.7 \text{ mg L}^{-1} \text{ O}_2$ ), temperature ( $30 \text{ }^\circ\text{C}$ ), and photoperiod (12 h lightness: 12 h dark) constants. In each sampling unit 0.3 g of the salt  $\text{L}^{-1}$  was added.

The water at the beginning of the experiment had: alkalinity (30.20 mg L<sup>-1</sup> of CaCO<sub>3</sub>), pH (7.0), calcium (6.0 mg L<sup>-1</sup>), hardness (17.0 mg L<sup>-1</sup>), silica (0.02 mg L<sup>-1</sup>) and magnesium (11.0 mg L<sup>-1</sup>). The soil samples (Table 1) were air-dried, homogenized, and sieved through a 2 mm mesh (sieve no. 10). Was added 1.2 liters of soil aquarium<sup>-1</sup>.

Table 1. Mean values and standard deviation of the soil composition.

Textural composition	Amount (%)
Sand	37.8 ± 0.42b
Silt	46.0 ± 0.00a
Clay	16.2 ± 0.42c
Sieve aperture (mm)	Retained amount (%)
2,00	11.6 ± 0.65c
1,00	13.6 ± 0.58c
0,50	17.4 ± 0.29b
0,250	19.0 ± 0.14b
0,106	26.7 ± 0.81a
<0,106	11.4 ± 0.82c
Physico-chemical characteristics	Mean values
pH	6.9 ± 0.51
Redox potential (mV)	242.3 ± 17.55
Electric conductivity (mS cm <sup>-1</sup> )	0.03 ± 0.02
Density (g L <sup>-1</sup> )	1.07 ± 0.02

Means followed by distinct letters, in the same sections of the columns, differ by the Tukey's test, 0.05 probability.

The experiment began with 375 specimens of Nile tilapia larvae weighing  $0.02 \pm 0.00$  g, measuring the standard length of  $1.12 \pm 0.08$  cm and total length of  $0.93 \pm 0.08$  cm, and subsequently distributed 15 animals per aquarium at a density of 1.5 larvaL<sup>-1</sup>. The animals were fed commercial powdered feed, with crude protein (min.) 550 g kg<sup>-1</sup>, ether extract (min.) 80 g kg<sup>-1</sup>, fibrous matter (max.) 30 g kg<sup>-1</sup>, mineral matter (max.) 160 g kg<sup>-1</sup>, calcium (max.) 30 g kg<sup>-1</sup>, phosphorus (min.) 14 g kg<sup>-1</sup> and moisture (max.) 100 g kg<sup>-1</sup>, according to the manufacturer's specifications, offering until apparent satiation, in three meals: 8, 12 and 16 h.

Twice a week (Monday and Thursday), the aquariums were cleaned by siphoning, and 20% of the volume was renewed and replaced with a specific stock solution for each treatment. Before feeding the fish, every fifteen days (1, 15, and 30 days), water samples were obtained from each aquarium to control their parameters. The parameters measured were: temperature (°C), pH, redox potential (mV), conductivity (mScm<sup>-1</sup>), salinity (‰), turbidity (NTU) using a HORIBA U10® measuring probe; alkalinity (mg L<sup>-1</sup>), hardness (mg L<sup>-1</sup>), calcium (mg L<sup>-1</sup>), magnesium (mg L<sup>-1</sup>)

using the titrimetric method; and the concentrations of ammonia ( $\text{mg L}^{-1}$ ), nitrite ( $\text{mg L}^{-1}$ ), nitrate ( $\text{mg L}^{-1}$ ) and silica ( $\text{mg L}^{-1}$ ) using the spectrophotometric method, as indicated by APHA<sup>13</sup>.

At the end of the experiment, measurements of total and standard length (cm) were taken with a digital caliper (Starret) with an accuracy of 0.02 mm and weight (g) with an analytical balance with an accuracy of 0.01 g when the number of individuals was also counted to calculate survival (%). From these records, weight gain (g) = (final weight - initial weight), biomass (g) (average weight x number of specimens per aquarium), biomass gain (g) = (final biomass - initial biomass), and Fulton's condition factor (K) [ $= 100 \times (\text{total length weight}^{-3})$ ], were calculated.

The data were subjected to normality tests (Shapiro-Wilk) and homoscedasticity. Survival data were transformed into arcsine for statistical analysis but were shown in percentages. The data were subjected to one-way ANOVA and Tukey's test at a significance level 0.05. For calculations, the R software was used.

## RESULTS

The parameters of growth and survival of Nile tilapia larvae are presented in Table 2. Total length, standard length, weight, weight gain, biomass, biomass gain, Fulton condition factor, and survival did not show differences between treatments.

The water quality parameters in the different treatments in the Nile tilapia larvae culture are shown in Tables 3, 4, and 5. The data on dissolved oxygen in the water, nitrite, nitrate, and magnesium did not differ significantly according to the products tested ( $p > 0.05$ ). The parameters pH, redox potential, electrical conductivity, turbidity, salinity, ammonia, alkalinity, hardness, calcium, and silica showed significant differences ( $p < 0.05$ ) between the liming products and the soil, as well as the absence of them.

Table 2. Mean values, standard deviation, and coefficient of variation from the performance of the Nile tilapia larvae subjected to different liming products for 30 days.

	Weight (g)	Total length (cm)	Standard length (cm)	Fulton condition factor
Water	3.63±0.22	2.86±0.23	0.73±0.19	15.83±2.86
Calcitic	3.54±0.08	2.72±0.12	0.59±0.09	17.66±1.90
Dolomitic	3.70±1.54	2.89±1.20	0.76±0.42	16.38±5.23
Soil	3.65±0.19	2.84±0.14	0.77±0.16	16.01±1.45
Silicate	3.61±0.16	2.83±0.13	0.67±0.10	15.92±1.73
CV (%)	7.76	8.73	30.13	19.56
	Weight gain (g)	Biomass (g)	Biomass gain (g)	Survival (%)
Water	3.61±0.22	50.23±9.53	49.93±9.53	92.0±16.0
Calcitic	3.52±0.08	51.05±2.17	50.75±2.17	96.0±3.26
Dolomitic	3.68±0.47	43.49±19.26	43.19±19.26	64.0±44.54
Soil	3.63±0.19	50.55±6.47	50.25±6.47	92.0±9.79
Silicate	3.59±0.16	51.37±5.06	51.37±5.06	94.6±7.77
CV (%)	7.8	22.19	22.32	20.92

In the same column, means did not differ, according to the Tukey test, at 0.05 probability.

The pH was higher ( $p < 0.05$ ) in the silicate and calcitic treatments than in the control, and the others did not differ from each other. For the redox potential, the dolomitic treatment was higher ( $p < 0.05$ ) than in the soil, and the others did not differ from each other. The electrical conductivity was higher ( $p < 0.05$ ) in the soil treatment than in the control, and the others did not differ from each other. The turbidity in the soil was higher ( $p < 0.05$ ) than in the other treatments.

Table 3. Mean values, standard deviation, and coefficient of variation of the water quality of Nile tilapia larvae culture subjected to different liming products for 30 days.

	pH	ORP (mV)	Conductivity (mS cm <sup>-1</sup> )	Turbidity (NTU)
Water	7.47 ±0.49c	190.4±47.47ab	0.20±0.01b	169.1±111.66b
Calcitic	7.97±0.28a	162.9±41.34ab	0.21±0.00ab	209.1±98.99b
Dolomitic	7.81±0.37ab	194.8±62.55a	0.21±0.00ab	163.7±136.89b
Soil	7.58±0.42bc	156.0±21.42b	0.35±0.02a	355.1±214.51a
Silicate	8.09±0.38a	170.6±49.12ab	0.28±0.00ab	164.0±186.91b
CV (%)	3.81	20.83	35.42	66.21

Means followed by different letters in the columns differ, using the Tukey test, at 0.05 probability. ORP= oxidation reduction potential.

Salinity was higher ( $p < 0.05$ ) in treatments that received liming products but with low values. Ammonia was higher ( $p < 0.05$ ) in the control treatment than in the treatment with silicate, and the others did not differ from each other. Nitrite and nitrate concentrations did not differ ( $p > 0.05$ ) between treatments.

Table 4. Mean values, standard deviation, and coefficient of variation of the water quality of Nile tilapia larvae culture subjected to different liming products for 30 days.

	DO (mg L <sup>-1</sup> )	Salinity (‰)	Ammonia (mg L <sup>-1</sup> )	Nitrite (mg L <sup>-1</sup> )	Nitrate (mg L <sup>-1</sup> )
Water	4.8±0.73a	0.00±0.00b	0.03±0.03a	0.16±0.27a	0.17±0.33a
Calcitic	4.7±0.66a	0.01±0.00ab	0.02±0.02ab	0.14±0.25a	0.18±0.25a
Dolomitic	6.6±4.75a	0.01±0.00a	0.02±0.03ab	0.09±0.16a	0.11±0.31a
Soil	4.7±1.05a	0.00±0.00b	0.02±0.02ab	0.07±0.18a	0.02±0.08a
Silicate	4.8±0.81a	0.01±0.00a	0.01±0.01b	0.02±0.06a	0.05±0.12a
CV (%)	45.74	39.47	65.17	192.31	187.25

Means followed by distinct letters in the column vary by Tukey's test at 0.05 probability. ns Not significant. \* Significant by Tukey's test at 0.05 probability. DO = Dissolved oxygen.

Alkalinity was higher ( $p < 0.05$ ) in the silicate treatment and did not differ among the others. Hardness was higher ( $p < 0.05$ ) in the calcitic, dolomitic, and silicate treatments to the control and soil. Calcium dissolved in water was higher ( $p < 0.05$ ) in the calcitic and silicate treatments than in the control and soil. Silica dissolved in water was higher ( $p < 0.05$ ) in silicate.

Table 5. Mean values, standard deviation, and coefficient of variation of the water quality of Nile tilapia larvae culture subjected to different liming products for 30 days.

	Alkalinity (mg L <sup>-1</sup> )	Hardness (mg L <sup>-1</sup> )	Calcium (mg L <sup>-1</sup> )	Magnesium (mg L <sup>-1</sup> )	Silica (mg L <sup>-1</sup> )
Water	48.80±17.84b	39.3±22.45b	27.6±20.09b	11.6±8.31a	0.01±0.00b
Calcitic	52.27±20.64b	84.74±27.13a	70.3±31.14a	11.6±10.28a	0.01±0.01b
Dolomitic	52.73±19.30b	69.3±31.21a	53.3±23.09ab	16.0±19.35a	0.01±0.00b
Soil	51.27±24.20b	37.0±14.36b	26.3±11.33b	10.6±7.27a	0.01±0.01b
Silicate	67.27±25.59a	90.7±39.48a	79.07±35.95a	14.3±13.16a	0.30±0.01a
CV (%)	22.54	21.24	22.65	80.96	29.95

Means followed by distinct letters in the column vary by Tukey's test at 0.05 probability. ns Not significant. \* Significant by Tukey's test at 0.05 probability.

## DISCUSSION

### Larval performance

Larval performance and survival did not differ according to the liming products. Although these products provided differences in water quality parameters, they remained within the range considered suitable for the cultivation of larvae<sup>14-16</sup> and juveniles<sup>17,18</sup> of the species, which explains the similarity in animal performance between the treatments tested.



## Water quality

Water quality is the most critical factor affecting the health and performance of fish in production systems. They depend on the water they live in, which makes it necessary to understand the water quality conditions required by fish<sup>19</sup>.

The temperature (30 °C) maintained by the heaters was similar between treatments and within the suitable range for the species. Studies report that the appropriate range varies from 20.2 to 31.7°C, with the ideal temperature being 26.0°C<sup>20</sup>, with the highest growth rates observed at 26 and 30 °C<sup>21</sup> and 28 and 34°C<sup>22</sup>.

Throughout the period, aeration maintained similar dissolved oxygen levels between treatments and within the range (5 to 6 mg L<sup>-1</sup>) considered adequate for the survival and growth of Nile tilapia<sup>23-25</sup> and for tropical species<sup>26</sup>.

The redox potential (ORP) showed averages between 156.0 and 194.8 mV, close to values considered adequate for tilapia cultivation in a biofloc system, 155.33 mV<sup>27</sup>. The redox potential values of this experiment are consistent with the oxygen concentrations, which were maintained at constant rates (> 4.7 mg L<sup>-1</sup>) and with the pH (7.47 to 8.09) associated with periodic cleaning to remove organic matter. Pond management influences the behavior of the redox potential, and knowing the potential profile can help understand the welfare and immunity of animals<sup>28</sup>. A high and positive redox potential generally indicates an oxidizing environment, rich in oxygen and more suitable for aquatic organisms, while an environment with negative values indicates reducing water<sup>27-29</sup>. Under high redox values, conditions are usually favorable for bacteria to decompose organic matter and contaminants more efficiently<sup>30</sup>. Several factors interact and interfere with the redox potential, such as the distribution in the water column, whether closer to the surface or the sediment-water interface, and factors such as the decomposition of organic matter and the metabolism of microorganisms, which decrease the potential, among others<sup>29</sup>. Therefore, the oxidation-reduction potential conditions were favorable to tilapia in all treatments due to this experiment's relatively homogeneous environments, with constant oxygenation, water renewal, and periodic cleaning in a reduced space.

Even though the initial phases of tilapia are more susceptible than adult specimens to changes in pH<sup>31,32</sup> the narrow pH range observed in this experiment was favorable to the well-being and development of the species, which can be measured by the survival values found.

The water in the aquariums without the addition of products and with soil at the bottom that did not receive liming products had a slight pH fluctuation, as in the other treatments, due to the alkalinity of their waters (48.80 and 51.27 mg L<sup>-1</sup>, respectively). The pH keeping occurs because alkalinity has a buffering effect that minimizes fluctuations in water pH, avoiding stress in fish, with the minimum values suggested for aquaculture being 20 mg L<sup>-1</sup> CaCO<sub>3</sub> of alkalinity<sup>6</sup> to 40 mg L<sup>-1</sup> CaCO<sub>3</sub><sup>33</sup>. Soil aquariums could present low pH and alkalinity, being susceptible to sudden changes in pH since the soil is usually a source of acidity due to aluminum ions, clay, and negatively charged organic matter particles, attracting cations to their surfaces, in addition to low concentrations of bicarbonate, carbonate, calcium, and magnesium<sup>34</sup>, which was not observed in this experiment, in which the pH of the water, in the various treatments, ranged from 7.5 (water) to 8.1 (silicate), being within the range considered suitable for the growth of larvae<sup>14,35,36</sup> and juveniles<sup>37</sup> of Nile tilapia.

The alkalinity of this experiment presented an adequate range with slight oscillation (48.80 to 67.27 mg L<sup>-1</sup>), which explains why no difference was observed in larval growth. The values were almost similar for all treatments except for silicate, which were slightly higher.

Although waters with higher alkalinity tend to have higher fish production due to the more significant buffering effect, with less fluctuation in pH<sup>4</sup>, the results depend on the alkalizing product<sup>8,9</sup> and the adequate concentration, for each species and stage of development<sup>31,38</sup>.

The values were similar to those found in the culture of Nile tilapia larvae, which grew more when subjected to 32 mg L<sup>-1</sup> of CaCO<sub>3</sub> than to 15 or 55 mg L<sup>-1</sup> of CaCO<sub>3</sub><sup>31</sup>. The alkalinity levels were also similar to those observed in the culture of tilapia juveniles in bioflocs in which sodium bicarbonate (NaHCO<sub>3</sub> 75.76 mg L<sup>-1</sup>), calcium carbonate (CaCO<sub>3</sub> 49.95 mg L<sup>-1</sup>) or calcium hydroxide (Ca(OH)<sub>2</sub> 54.58 mg L<sup>-1</sup>) were used, in which the juveniles presented greater weight when subjected to the highest alkalinity (75.76 mg L<sup>-1</sup>), and worse in the lowest (49.95 mg L<sup>-1</sup>), a difference that was associated with a change in management due to the characteristics of the alkalizing agents<sup>9</sup>. The survival (91.0%) of the fish was similar to that found for Nile tilapia (88.3%) subjected to different alkalinities (9.2, 15.5, 30.8, 73.0, and 90.4 mg L<sup>-1</sup>) obtained by other products (CaCO<sub>3</sub>, Na<sub>2</sub>CO<sub>3</sub> or CaSO<sub>4</sub>) where the increase in alkalinity by CaSO<sub>4</sub> provided a negative effect<sup>39</sup>. The values observed in this experiment, adding the values of the studies above, indicate that the alkalinity was within the range suitable for the growth of Nile tilapia larvae.

Despite the lower hardness levels observed in aquariums without products (control) and in soil, the hardness range in this experiment was within that considered adequate for the species<sup>39-41</sup>, which generated similar animal performance between treatments. Water hardness increases the antioxidant response, reduces the toxicity of some substances, the loss of ions, buffering the environment and favoring fish development<sup>42-45</sup>. Nile tilapia juveniles perform better when total hardness is greater than 20 mg CaCO<sub>3</sub> L<sup>-1</sup><sup>39</sup>, and when water hardness is less than 25 mg L<sup>-1</sup>, limestone should be applied<sup>46</sup>. Since the water used in the experiment initially had 17 mg L<sup>-1</sup> and increased in the control treatment, which was only water, calcium in the feed helped to increase hardness. The initial hardness required the application of the alkalizing agent. Increased hardness and decreased alkalinity in closed aquaculture systems have been reported<sup>47</sup>.

Regarding calcium, a higher amount occurred in the treatments that received liming products, similar to that observed by Antonangelo et al.<sup>48</sup>. However, even the control and soil group showed an increase in the amount of calcium dissolved in the water compared to the first day of collection due to the calcium in the feed offered to the fish, as observed in the cultivation of cachama blanca, *Piaractus brachypomus* in a recirculation system, where hardness increased<sup>49</sup>. Magnesium, on the other hand, was not added enough to provide a significant difference, unlike that observed by Antonangelo et al.<sup>48</sup>.

Calcium and magnesium ions are essential for water hardness and alkalinity<sup>6</sup> and ionic regulation in freshwater fish<sup>39</sup>. Aquatic animals have unique physiological mechanisms to absorb and retain minerals, such as calcium, magnesium, and phosphorus, through their diets and directly from water, through gill and skin absorption, with excessive intake or deficiency being harmful to fish<sup>50,51</sup>. Magnesium deficiency causes skeletal deformities, cardiovascular diseases, and metabolic syndrome<sup>52</sup>, such as nephrocalcinosis<sup>51</sup>, damage to the structural integrity of the intestine, suppressing fish growth<sup>53</sup>. Calcium deficiency compromises skeletal formation<sup>54</sup>, resulting in fish with deformities<sup>50</sup>.

Excess is also a problem, as high levels of calcium (31.0 g kg<sup>-1</sup> of feed) in the diet of Japanese sea bass, *Lateolabrax japonicus*, significantly reduced weight, weight gain, feeding rate, specific growth rate, whole-body and muscle protein and lipid contents, as well as serum Ca concentration and alkaline phosphatase activity. There was even a significant reduction in the vertebral contents of Ca, P, Zn, Fe, and Mn and the contents of Ca, P, Mg, and Mn in scales, leading to a higher rate of fish deformation<sup>50</sup>. It is also worth noting that magnesium helps to activate

vitamin D, which helps to regulate calcium and phosphate homeostasis, which influences bone growth and maintenance<sup>52</sup>.

As no deformities were observed and growth was equal between treatments and within expectations, calcium and magnesium levels were adequate for tilapia larvae.

The higher concentration of silica in water with added silicate was expected, as also observed by Emerenciano et al.<sup>55</sup>. Silica is the second most abundant component in the Earth's crust, making up the silicate of rocks that dissolve in natural waters<sup>56</sup>. However, due to the low solubility of this mineral, these waters generally have low alkalinity and total hardness, both in surface waters and aquifers composed of sand and/or silicate minerals<sup>7</sup>. However, when in the form of calcium silicate ( $\text{CaSiO}_3$ ) or sodium silicate ( $\text{Na}_2\text{SiO}_3$ ), the solubility is sufficient for this material to be used for liming, neutralizing the hydrogen ion, in addition to serving as a source of silica for diatoms<sup>7,56</sup>, which increase their population, improving shrimp growth performance, inhibiting pathogenic vibrios, increasing the profit of the activity<sup>55</sup>.

Calcium-magnesium silicate was promising as a promoter of a stable environment for tilapia larvae culture, as it allowed suitable water quality parameters, in addition to the development and survival being similar to those of tilapia larvae grown in waters with calcium carbonate and calcium and magnesium carbonate, known to be suitable for fish farming<sup>7</sup>. The results of the larvae in water with silicate were also similar to those kept in water with soil, an element that makes up the natural environment of tilapia. Menezes et al.<sup>57</sup> obtained similar results in the yield and survival of tilapia juveniles subjected to water with the addition of calcium silicate, dolomitic limestone and calcitic limestone, as well as de Souza et al.<sup>8</sup> obtained similar results in the yield and survival of jundiá *Rhamdia quellen* juveniles subjected to water with the addition of sodium silicate, sodium bicarbonate and calcium carbonate.

Agronomic studies also reinforce the findings regarding water quality in this experiment, where calcium and magnesium silicate significantly increased pH, calcium, magnesium, and silicon, reducing the elements that promote acidity<sup>48</sup>. Other results evaluating calcium and magnesium silicate prove the high power of neutralizing soil acidity when compared to dolomitic limestone<sup>58</sup> and that calcium silicate is 6.8 times more soluble than calcium carbonate, favoring the correction of acidity<sup>59</sup>.

The electrical conductivity was within the acceptable range,  $1000 \mu\text{S cm}^{-1}$ , for fish farming<sup>6</sup>. The conductivity of the water in aquariums with soil was significantly higher than that of aquariums with only water. The others were similar to all.

Tank soils interact with water, altering its characteristics<sup>60</sup>. Soil conductivity results from its composition, with soils richer in clay and smaller particles with higher conductivity than sandy soils<sup>61</sup>. Therefore, the soil's presence explains the water's higher conductivity.

However, despite increasing the electrical conductivity of the water<sup>62,63</sup>, the different liming products did not increase the conductivity compared with the control treatment (water only). High conductivities can also be related to a large amount of organic matter in the water, usually from the feed<sup>64</sup>. Waters rich in phosphate, among other nutrients, generally have a higher conductivity, which could make the electrical conductivity of the water an indicator of eutrophication<sup>61</sup>. However, the amount of nutrients added by the feed was similar between treatments, justifying the high conductivity values and the similarity of the results.

The turbidity of the soil treatment was higher than that of the others, which the suspension of sediment particles can explain. Suspended clay is responsible for much of the turbidity of the water in fish ponds<sup>65</sup>.

Turbidity has been considered a problem in the cultivation of Nile tilapia juveniles<sup>9,65,66</sup> when it affects fish development<sup>9,66</sup>, and even when it does not interfere<sup>65</sup>. The increase in turbidity can also be caused by liming products added to the environment<sup>9,67</sup>, although it did not increase turbidity in this experiment. Although tilapia do not suspend much clay from the bottom of excavated tanks<sup>66</sup>, they increase turbidity by suspending particles from the bottom of the tank in this intensive system due to the fish swimming in environments with reduced space.

Total ammonia concentrations were low and within the comfort range for cultivating tilapia larvae<sup>14,36</sup>.

Considering that Nile tilapia larvae did not have their growth and survival compromised when subjected for 80 days after hatching to a concentration of  $0.25 \text{ mg L}^{-1}$  of non-ionized ammonia, becoming affected at  $0.45 \text{ mg L}^{-1}$ <sup>68</sup>, and the average  $\text{LC}_{50}$  value at 48 h was  $1,009 \text{ mg L}^{-1}$  for the larvae<sup>69</sup>, the ammonia concentration was within the adequate range for Nile tilapia larvae.

As with ammonia, nitrite and nitrate concentrations were similar to the values considered adequate for raising Nile tilapia larvae<sup>14,36</sup>, and were below the concentrations considered safe

limits for fish farming, which are  $0.5 \text{ mg L}^{-1}$  and  $4.5\text{-}5.0 \text{ mg L}^{-1}$  of nitrite and nitrate, respectively<sup>70</sup>,<sup>71</sup> or even of the ideal concentrations of  $0.3 \text{ mg L}^{-1}$  and  $25.0 \text{ mg L}^{-1}$  of nitrite and nitrate, respectively<sup>72</sup>.

Although these limit values exist, the nitrite level considered comfortable varies with the species, stage of development, and interactions with other water parameters. For juveniles of the yam *Cichlasoma facetum*, a cichlid like tilapia, the safety limit is  $7.0 \text{ mg L}^{-1}$ <sup>73</sup>, while for juveniles of *Rhamdia quelen*, the adequate limit that does not compromise growth and survival is  $1.19 \text{ mg L}^{-1}$ <sup>74</sup>. Rainbow trout gets stressed by  $0.15 \text{ mg L}^{-1}$  of nitrite and die by  $0.55 \text{ mg L}^{-1}$ . Channel catfish are more resistant to nitrite, starting to die at  $29 \text{ mg L}^{-1}$ . However, water should be checked whenever  $0.1 \text{ mg L}^{-1}$  or more of nitrite is present<sup>73</sup>. Although nitrite values in the aquariums with only water ( $0.16 \text{ mg L}^{-1}$ ) and in the calcitic limestone (calcium carbonate –  $0.14 \text{ mg L}^{-1}$ ) were above  $0.1 \text{ mg L}^{-1}$ , they were adequate for the species.

The low values of nitrogen compounds explain why the alkalinities did not change much. The nitrification process by bacteria reduces alkalinity by releasing  $\text{H}^+$  ions into the water<sup>47</sup>, while the oxidation of  $1 \text{ mg L}^{-1}$  of ammonia reduces alkalinity by  $7.14 \text{ mg L}^{-1}$ <sup>7</sup>. Therefore, the low nitrogen compound concentrations resulted in low alkalinity consumption, which remained within a range suitable for Nile tilapia larvae.

The salinity variation was slight and remained within the range considered freshwater when the salinity is less than  $1\text{‰}$ <sup>75</sup>. The early life stages of Nile tilapia are euryhaline. Although low salinities negatively affect hatching rates, survival, and yolk sac absorption, larvae six days after hatching tolerate salinities of up to  $20\text{‰}$ <sup>76</sup>. Since Nile tilapia tolerate salinities from  $7\text{‰}$ <sup>77</sup> to  $22.5\text{‰}$ <sup>78</sup> and can be cultured in waters with up to  $30\text{‰}$ , if properly acclimated and fed<sup>79</sup>, the salinities were adequate for larval development.

## CONCLUSION

Calcium-magnesium silicate is a viable and recommended alternative for liming, as it presents results equivalent to calcitic and dolomitic limestone, traditional salts for this practice, which confirms its importance in maintaining water quality and fish performance. Although the soil has a buffering capacity in the water, it is less effective than other products and deserves further study.

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## REFERENCES

1. Verdegem M, Buschmann AH, Latt UW, Dalsgaard AJ, Lovatelli A. The contribution of aquaculture systems to global aquaculture production. *Journal of the World Aquaculture Society*. 2023; 54(2): 206-250. <https://doi.org/10.1111/jwas.12963>.
2. Laktuka K, Kalnbalkite A, Sniega L, Logins K, Lauka D. Towards the sustainable intensification of aquaculture: Exploring possible ways forward. *Sustainability*. 2023; 15(24): 16952. <https://doi.org/10.3390/su152416952>.
3. Jafari L, Montjouridès MA, Hosfeld CD, Attramadal K, Fivelstad S, Dahle H. Biofilter and degasser performance at different alkalinity levels in a brackish water pilot scale recirculating aquaculture system (RAS) for post-smolt Atlantic salmon. *Aquacultural Engineering*. 2024; 106: 102407. <https://doi.org/10.1016/j.aquaeng.2024.102407>.
4. Boyd, CE, Tucker, CS, Somridhivej, B Alkalinity and hardness: critical but elusive concepts in aquaculture. *Journal of the World Aquaculture Society*, 2016; 47(1): 6-41. <https://doi.org/10.1111/jwas.12241>.
5. Pedreira MM Calagem na manutenção da qualidade da água na aquicultura. In: Membrive CMB, Bernardes EM, Rosas FS, Fonseca R (eds) *IMAST 2023: IV International Meeting of Agrarian Science and Technology*. FCAT, Dracena; Ladri Marília, SP: UNESP; 2023. cap. 10, p. 154–171. [https://www.dracena.unesp.br/Home/Servicos/Biblioteca/livrosfcats/livrodig\\_imast\\_2023.pdf](https://www.dracena.unesp.br/Home/Servicos/Biblioteca/livrosfcats/livrodig_imast_2023.pdf).
6. Boyd CE, Tucker CS. Handbook for aquaculture water quality. *Handbook for Aquaculture Water Quality*, 2014; 439p.
7. Boyd CE Tucker CS *Pond aquaculture water quality management*. New York: Springer Science & Business Media; 2012. 700p.
8. de Souza ER, Ferreira TA, Pelli A, Moreira NF, Verardo LL, Pedreira MM. Alkalizing potentials for recirculating systems with clear water in the *Rhamdia quelen* juvenile cultivation. *Aquaculture International*. 2024; 1-17. <https://doi.org/10.1007/s10499-024-01573-6>.
9. Martins GB, Tarouco F, Rosa CE, Robaldo RB The utilization of sodium bicarbonate, calcium carbonate or hydroxide in biofloc system: water quality, growth performance and oxidative stress

of Nile tilapia (*Oreochromis niloticus*). *Aquaculture*. 2017; 468: 10–17. <https://doi.org/10.1016/j.aquaculture.2016.09.046>.

10. Boyd, CE. The importance of liming materials in aquaculture: Calcium carbonate, magnesium carbonate essential in management of production ponds. *Global Seafood Advocate*. 2016; Available at: ([globalseafood.org](http://globalseafood.org)), accessed at: 09 august 2024.

11. Avnimelech Y, Ritvo G. Shrimp and fish pond soils: processes and management. *Aquaculture*. 2003; 220(1-4): 549-567. [https://doi.org/10.1016/S0044-8486\(02\)00641-5](https://doi.org/10.1016/S0044-8486(02)00641-5).

12. Dong SL, Li L. Sediment and remediation of aquaculture ponds. In: Dong SL, Tian XL, Gao QF, Dong YW. (eds) *Aquaculture Ecology*. Singapore: Springer; 2023. [https://doi.org/10.1007/978-981-19-5486-3\\_8](https://doi.org/10.1007/978-981-19-5486-3_8).

13. APHA - American Public Health Association. Standard methods for the examination of water and wastewater. 22nd. Rice, E.W.; Baird, R.B.; Eaton, A.D.; Clesceri, L.S. Washington, D.C.: American Public Health Association, American Water Works Association, Water Environment Federation. 2012. 1496p.

14. Silva, ED, Pedreira, MM., Dias, M.F, Tessitore ADA, Ferreira TA. Larvae of Nile tilapia lines subject to feeding frequencies under low temperature. *Revista Brasileira de Saúde e Produção Animal*. 2017; 18(1):193-203. <https://doi.org/10.1590/S1519-99402017000100018>.

15. Li J, Liua G, Li C, Deng Y, Tadda MA, Lan L, Zhu S, Liu D. Effects of different solid carbon sources on water quality, biofloc quality and gut microbiota of Nile tilapia (*Oreochromis niloticus*) larvae. *Aquaculture*. 2018; 495: 919-931. <https://doi.org/10.1016/j.aquaculture.2018.06.078>

16. Ali A, Moustafa YT, El-Said S. Evaluating the influence of different water sources on water quality, survival and growth rates of Nile tilapia (*Oreochromis niloticus*) larvae in tilapia hatcheries. *Egyptian Journal for Aquaculture*. 2020; 10(1): 45-64. <https://doi.org/10.21608/eja.2020.25601.1018>.

17. Martins GB, da Rosa CE, Tarouco FDM, Robaldo RB. Growth, water quality and oxidative stress of Nile tilapia *Oreochromis niloticus* (L.) in biofloc technology system at different pH. *Aquaculture Research*. 2019; 50(4): 1030-1039. <https://doi.org/10.1111/are.13975>.

18. Susitharan V, Krishnan S, Kumar P, Sukhdhane K, Kala AS, Rani AB. Mineral supplementation in biofloc influences growth and haemato-biochemical indices of Genetically Improved Farmed Tilapia reared in inland saline ground water. *Aquacultural Engineering*. 2024; 104. 102386. <https://doi.org/10.1016/j.aquaeng.2023.102386>.

19. Akhter F, Siddiquei HR, Alahi MEE, Mukhopadhyay SC. Recent advancement of the sensors for monitoring the water quality parameters in smart fisheries farming. *Computers*. 2021; 10(3): 26. <https://doi.org/10.3390/computers10030026>.



20. Leonard JN, Skov PV. Capacity for thermal adaptation in Nile tilapia (*Oreochromis niloticus*): Effects on oxygen uptake and ventilation. *Journal of Thermal Biology*. 2022; 105: 103206. <https://doi.org/10.1016/j.jtherbio.2022.103206>.
21. Hamed SA, Abou-Elnaga A, Salah AS, Abdel-Hay AHM, Zayed MM, Soliman T, Mohamed RA. Effect of water temperature, feeding frequency, and protein percent in the diet on water quality, growth and behavior of Nile tilapia *Oreochromis niloticus* (Linnaeus, 1758). *Journal of Applied Ichthyology*. 2021; 37(3): 462-473. <https://doi.org/10.1111/jai.14193>.
22. Zeng NN, Jiang M, Wen H, Liu W, Wu F, Tian J, ... Guo ZB Effects of water temperatures and dietary protein levels on growth, body composition and blood biochemistry of juvenile GIFT tilapia (*Oreochromis niloticus*). *Aquaculture Nutrition*. 2021; 27(1): 240-251. <https://doi.org/10.1111/anu.13181>.
23. Santos VB, Mareco EA, Silva MDP. Growth curves of Nile tilapia (*Oreochromis niloticus*) strains cultivated at different temperatures. *Acta Scientiarum. Animal Sciences*. 2013; 35(3): 235–242.
24. Abdel-Tawwab M, Hagraas AE, Elbaghdady, HAM, Monier MN. Effects of dissolved oxygen and fish size on Nile tilapia, *Oreochromis niloticus* (L.): growth performance, whole-body composition, and innate immunity. *Aquaculture International*. 2015; 23: 1261-1274. <https://doi.org/10.1007/s10499-015-9882-y>.
25. Khanjani MH, Sharifinia M. Production of Nile tilapia *Oreochromis niloticus* reared in a limited water exchange system: The effect of different light levels. *Aquaculture*. 2021; 542: 736912. <https://doi.org/10.1016/j.aquaculture.2021.736912>.
26. Mallya YJ The effects of dissolved oxygen on fish growth in aquaculture. Reykjavík, Ísland: Kingolwira National Fish Farm Center UNU-Fisheries Training Programme, 30 pp. 2007.
27. Magondu E W, Verdegem MCJ, Nyakeya K, Mokaya M. Production of aerobic, anaerobic and anoxic bioflocs from tilapia sludge. *International Journal of Fisheries and Aquatic Studies*. 2015; 2(5): 347-352.
28. Lehmann M, Vinatea L. Redox potential in freshwater and seawater culture ponds: determination methodology behavior. *Boletim do Instituto de Pesca*. 2008; 34: 131-140.
29. Boyd, CE. 3rd Water quality: an introduction. Cham: Springer Nature; 2020. 452 p. <https://doi.org/10.1007/978-3-030-23335-8>.
30. Birchenough S, Parker R, Mcmanus E, Barry J. Combining bioturbation and redox metrics: potential tools for assessing seabed function. *Ecological Indicators*. 2012; 12(1): 8–16. <https://doi.org/10.1016/j.ecolind.2011.03.015>.
31. Rojas NET, Rocha O. Influência da alcalinidade da água sobre o crescimento de larvas de tilápia do Nilo (*Oreochromis niloticus* Linnaeus, 1758 Perciformes, Cichlidae). *Acta Scientiarum. Biological Sciences*. 2004; 26(2): 163-167.

32. Cavalcante DH, Poliato AS, Ribeiro DC, Magalhães FB, Sá MVC. Effects of CaCO<sub>3</sub> liming on water quality and growth performance of fingerlings of Nile tilapia, *Oreochromis niloticus*. *Acta Scientiarum. Animal Sciences*. 2009; 31: 327-333.
33. Moro GV, Torati LS, Luiz DDB, Matos FD Monitoramento e manejo da qualidade da água em pisciculturas. In: Rodrigues APO, Lim AF, Alves AL, Rosa DK, Torati LS, dos Santos VRV (eds) *Piscicultura de água doce: multiplicando conhecimentos*. Brasília – DF: Embrapa; 2013. pp.141–169. <https://ainfo.cnptia.embrapa.br/digital/bitstream/doc/1083545/1/cap.5.pdf>. Accessed 28 Ago 2024.
34. Boyd CE, Wood CW, Thunjai T. *Aquaculture pond bottom soil quality management*. Pond Soils. Oregon State University, Corvallis, Oregon, USA; 2002.
35. Ekassari J, Rivandi DR, Firdausi AP, Surawidjaja EH, Zairin JRM, Bossier P, de Schryver P. Biofloc technology positively affects Nile tilapia (*Oreochromis niloticus*) larvae performance. *Aquaculture*. 2015; 441: 72-77. <https://doi.org/10.1016/j.aquaculture.2015.02.019>.
36. Pedreira MM, Tessitore AJA, Pires AV, Silva MA, Schorer M. Substrates for biofilter in recirculating system in Nile tilapia larviculture production. *Revista Brasileira de Saúde e Produção Animal*. 2016; 17(3): 553-560. <https://dx.doi.org/10.1590/S1519-99402016000300020>.
37. El-Sherif MS, El-Feky AMI. Performance of Nile tilapia (*Oreochromis niloticus*) fingerlings. I. Effect of pH. *International Journal of Agriculture and Biology*, 2009; 11(3): 297-300.
38. Ueno-Fukura M, Corredor-Ruiz JS, Jiménez-Ojeda YK, Collazos-Lasso LF. Usage of alkalizers in the nursery culture of *Piaractus brachypomus* with Biofloc technology-BFT. *Aquaculture, Aquarium, Conservation & Legislation*. 2019; 12(4): 989-995.
39. Cavalcante DH, Silva SR, Pinheiro PD, Akao MMF, Sá MVC. Single or paired increase of total alkalinity and hardness of water for cultivation of Nile tilapia juveniles, *Oreochromis niloticus*. *Acta Scientiarum. Technology*. 2012; 32(2): 177–183.
40. Cardoso Filho R, Campeche DF, Paulino RV. Tilápia em reservatório de água para irrigação e avaliação da qualidade da água. *Revista Brasileira de Ciências Agrárias*. 2010; 5(1): 117-122. <https://doi.org/10.5039/agraria.v5i1a669>.
41. Bart AN, Prasad B, Thakur DP. Effects of incubation water hardness and salinity on egg hatch and fry survival of Nile tilapia *Oreochromis niloticus* (Linnaeus). *Aquaculture Research*. 2012; 44(7): 1085–1092. doi:10.1111/j.1365-2109.2012.03113.x.
42. Baldisserotto B. Water pH and hardness affect growth of freshwater teleosts. *Revista Brasileira de Zootecnia*. 2011; 40(supl. especial): 138-144.
43. Parra JEG, Baldisserotto B. Effect of water pH and hardness on survival and growth of freshwater teleosts. In: *Fish osmoregulation*. Boca Raton, Florida: CRC Press; 2019. Cap. 3, p. 135-150.

44. Copatti CE, Baldisserotto B, Souza CDF, Monserrat JM, Garcia L. Water pH and hardness alter ATPases and oxidative stress in the gills and kidney of pacu (*Piaractus mesopotamicus*). *Neotropical Ichthyology*. 2019; 17(4): e190032. <https://doi.org/10.1590/1982-0224-20190032>.
45. Choi CY, Kim MJ, Song JA, Kho KH. Water hardness improves the antioxidant response of zinc-exposed goldfish (*Carassius auratus*). *Biology*. 2023; 12(2): 289. <https://doi.org/10.3390/biology12020289>.
46. Burtle GJ. Pond fertilization and liming in Georgia. UGA Extension Bulletin 867: 1-7. 2015.
47. Cavalcante DH, Caldini NN, Silva JLS, Lima FRS, Sá MVC. Imbalances in the hardness/alkalinity ratio of water and Nile tilapia's growth performance. *Acta Scientiarum*, 2014; 36(1): 49-54, <https://doi.org/10.4025/actascitechnol.v36i1.18995>.
48. Antonangelo JA, Neto JF, Crusciol CAC, Zhang H, Alleoni LRF. Lime and calcium-magnesium silicate cause chemical attributes stratification in no-till fields. *Soil and Tillage Research*. 2022; 224: 105522. <https://doi.org/10.1016/j.still.2022.105522>.
49. Poleo G, Aranbarrio JV, Mendoza L, Romero O. Cultivo de cachama blanca en altas densidades y en dos sistemas cerrados. *Pesquisa Agropecuária Brasileira*. 2011; 46(4): 429-437. <https://doi.org/10.1590/S0100-204X2011000400013>.
50. Song JY, Zhang CX, Wang L, Song K, Hu SC Zhang L. Effects of dietary calcium levels on growth and tissue mineralization in Japanese seabass, *Lateolabrax japonicus*. *Aquaculture Nutrition*. 2017; 23(3): 637-648. <https://doi.org/10.1111/anu.12431>.
51. Lall SP Kaushik SJ. Nutrition and metabolism of minerals in fish. *Animals*. 2021; 11(09): 2711. <https://doi.org/10.3390/ani11092711>.
52. Uwitonze AM, Razzaque MS. Role of magnesium in vitamin D activation and function. *Journal of Osteopathic Medicine*. 2018; 118(3): 181-189. <https://doi.org/10.7556/jaoa.2018.037>.
53. Wei SP., Jiang, WD., Wu, P., Liu1, Y., Zeng, Y.Y., Jiang, J., Kuang, S.Y., Tang, L., Zhang, Y.A., Zhou, X.Q. & Feng, L. (2018). Dietary magnesium deficiency impaired intestinal structural integrity in grass carp (*Ctenopharyngodon idella*). *Scientific Reports*. 8, 12705 <https://doi.org/10.1038/s41598-018-30485-8>.
54. Liu Y, Liu YN, Tian XC, Liu HP, Wen B, Wang N, Gao JZ, Chen ZZ. Growth and tissue calcium and phosphorus deposition of juvenile discus fish (*Symphysodon haraldi*) fed with graded levels of calcium and phosphorus. *Aquaculture*. 2021; 541: 736755. <https://doi.org/10.1016/j.aquaculture.2021.736755>.
55. Emerenciano MG, Arnold S, Perrin T. Sodium metasilicate supplementation in culture water on growth performance, water quality and economics of indoor commercial-scale biofloc-based *Litopenaeus vannamei* culture. *Aquaculture*. 2022; 560: 738566. <https://doi.org/10.1016/j.aquaculture.2022.738566>.

56. Boyd CE. Silicon, diatoms in aquaculture. *Global Aquaculture Advocate*. 2014; 17: 38-39.
57. Menezes WF, Souza ER, Pedreira RSF, Amorim MPS, Schorer M, dos Santos JCE, Pelli A, Pedreira MM. Calcium silicate and soil in the intensive cultivation of Nile tilapia. *Acta Biologica Brasiliensia*. 2023; 6(2): 43-62. <https://doi.org/10.18554/acbiobras.v6i2.7273>.
58. Calonego JC, Mora VS, Santos CH, de Oliveira L. Calagem e silicatagem em solo incubado com diferentes umidades. *Colloquium Agrariae*. 2012; 8(2): 46-56. <https://doi.org/10.5747/ca.2012.v08.n1.a078>.
59. de Lima Filho, OF, da Silva CJ Avaliação agrônômica do silicato de cálcio e magnésio granulado na cultura da cana-de-açúcar. 2017.
60. Saraswathy R, Muralidhar M, Sanjoy D, Kumararaja P, Suvana S, Lalitha N, Katneni VK, Nagavel A, Vijayan KK. Changes in soil and water quality at sediment–water interface of *Penaeus vannamei* culture pond at varying salinities. *Aquaculture Research*. 2019; 50(4): 1096-1106. <https://doi.org/10.1111/are.13984>.
61. Heiniger RW, McBride RG, Clay DE. Using soil electrical conductivity to improve nutrient management. *Agronomy Journal*. 2003; 95(3): 508-519. <https://doi.org/10.2134/agronj2003.5080>.
62. Nobre GR, Lima GS, Gheyi HR, Soares LAA, Silva AO. Crescimento, consumo e eficiência do uso da água pela mamoneira sob estresse salino e nitrogênio. *Revista Caatinga*. 2014; 27(2): 148 – 158.
63. Pieroni S, Olier BS, Lima IR, Sanches IM, Kuhnen VV, Sanches EG. Can use of substrates affect water quality in aquatic organism culture?. *Aquaculture International*. 2021; 29(4): 1771-1783. <https://doi.org/10.1007/s10499-021-00718-1>.
64. Akkoyunlu A Akiner ME. Pollution evaluation in streams using water quality indices: A case study from Turkey's Sapanca Lake Basin. *Ecological Indicators*. 2012; 18: 201-211. <https://doi.org/10.1016/j.ecolind.2011.12.018>.
65. Yi Y, Lin CK, Diana JS. Techniques to mitigate clay turbidity problems in fertilized earthen fish ponds. *Aquacultural Engineering*. 2003; 27(1), 39-51. [https://doi.org/10.1016/S0144-8609\(02\)00039-0](https://doi.org/10.1016/S0144-8609(02)00039-0).
66. Wing JDB, Champneys TS, Ioannou CC. The impact of turbidity on foraging and risk taking in the invasive Nile tilapia (*Oreochromis niloticus*) and a threatened native cichlid (*Oreochromis amphimelas*). *Behavioral Ecology and Sociobiology*. 2001; 75(3): 49. <https://doi.org/10.1007/s00265-021-02984-8>.
67. Helfrich LA, Neves RJ, Parkhurst JA. Liming acidified lakes and ponds. Publication 420-254. Petersburg: Virginia Cooperative Extension; 2001.

68. El-Greisy ZAEB, Ahmed NAM. Effect of prolonged ammonia toxicity on fertilized eggs, hatchability and size of newly hatched larvae of Nile tilapia, *Oreochromis niloticus*. Egyptian journal of aquatic research. 2016; 42(2): 215-222. <https://doi.org/10.1016/j.ejar.2016.04.001>.
69. Benli AÇK, Köksal G. The Acute Toxicity of Ammonia on Tilapia (*Oreochromis niloticus* L.) Larvae and Fingerlings. Turkish Journal of Veterinary & Animal Sciences. 2005; 29(2): 339-344. Article 23. Available at: <https://journals.tubitak.gov.tr/veterinary/vol29/iss2/23>.
70. Sipaúba-Tavares LHS. Limnologia aplicada à aquicultura. Jaboticabal – SP: FUNEP, 1995.
71. Santhosh B, Singh NP. Guidelines for water quality management for fish culture in Tripura, ICAR Research Complex for NEH Region, Tripura Center, Lembucherra-799210, Tripura (west). Publication no. 29, 2007.
72. Kubitzka F. Qualidade da água no SARs–monitoramento e correção. Panorama da Aquicultura. 2022; 31: 14-23.
73. Piedras SRN, Oliveira JLR, Moraes PRR, Bager A. Toxicidade aguda da amônia não ionizada e do nitrito em alevinos de *Cichlasoma facetum* (Jenyns, 1842). Ciência e agrotecnologia. 2006; 30: 1008-1012. <https://doi.org/10.1590/S1413-70542006000500027>.
74. Lima, RLD, Braun N, Kochhann D, Lazzari R, Radünz Neto J, Moraes BS, Loro V, Baldisserotto B. Survival, growth and metabolic parameters of silver catfish, *Rhamdia quelen*, juveniles exposed to different waterborne nitrite levels. Neotropical Ichthyology. 2011; 9(1):147-152. <https://doi.org/10.1590/S1679-62252011005000004>.
75. USGS Saline Water and Salinity. 2018. Disponível em: 15 setembro de 2024, Acessado em: Saline Water and Salinity | U.S. Geological Survey ([usgs.gov](https://www.usgs.gov)).
76. Fridman S, Bron J, Rana K. Influence of salinity on embryogenesis, survival, growth and oxygen consumption in embryos and yolk-sac larvae of the Nile tilapia. Aquaculture. 2012; 334: 182–190. doi:10.1016/j.aquaculture.2011.12.034.
77. de Azevedo RV, de Oliveira KF, Flores-Lopes F, Teixeira-Lanna EA, Takishita SS, Tavares-Braga LG Responses of Nile tilapia to different levels of water salinity. Latin American journal of aquatic research. 2015; 43(5): 828-835.
78. Kamal AHMM, Mair GC. Salinity tolerance in superior genotypes of tilapia, *Oreochromis niloticus*, *Oreochromis mossambicus* and their hybrids. Aquaculture. 2005; 247(1-4):189-201. <https://doi.org/10.1016/j.aquaculture.2005.02.008>.
79. Mirera DO, Okemwa D. Salinity tolerance of Nile tilapia (*Oreochromis niloticus*) to seawater and growth responses to different feeds and culture systems. Western Indian Ocean Journal of Marine Science. 2023; 22(2): 75-85. 10.4314/wiojms.v22i2.6.